Histological, Histochemical and Ultrastructural Studies on the Kidney of Rats After Administration of Monosodium Glutamate

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الملخص

دراسات هستولوجية وهستوكيميائية ودقيقة علي كلية الفئران بعد معالجتها بجلوتاميد أحادي الصوديوم

يُظهر هذا البحث تأثير إعطاء تركيزين مختلفين [(2)و (3) مجم/جمولزن جسم] من جلوتاميد أحادي الصوديوم لمدة 21, 45 يوم علي التوالي على التركيب النسيجي للكلى في ذكور الفئران البالغة . وقد أظهرت الفحوصات النسيجية لمجاميع الفئر ان التي عوملت بتركيز 2مجم/جم/وزن الجسم بداية فقدان جزئي للحافة الفرشاتية المكونة الأنيبيبات الملتفة الدانية.حدوث تحلل للطبقة الطلائية المكونة للأنابيب البولية التي ظهر فيها مجموعة من الخلايا الالتهابية . كذلك تَم ملاحظة تحلل جزئي وكلّي لأنوية الذي ظهر فيها مجموعة من الخلايا الالتهابية . كذلك تَم ملاحظة تحلل جزئي وكلّي لأنوية الخلايا المكونة لبعض نلك الأنابيب البولية. أما إعطاء جرعة 3مجم/جم/وزن الجسم ، فقد أظهر الفحص النسيجي ظهور تغيرات مرضية خطيرة تتمثل في تضخم الشعير ات الدموية المكونة لشعيرات الكبة وتكون حويصلات دهنية بها . وقد حدثت تشوهات في التركيب البنائي للخلايا المكونة للأنابيب البولية وقد حدثت التي تكونت في بعض خلايا الكبة التي أظهرت الفحوصات أيضاً سمك فـي الطبقـ المكونة للشعيرات الدموية في بعض الكبيبات الولية وحدوث نزيف داخل الفصيصات التي تكونت في بعض خلايا الكبة التي أظهرت الفحوصات أيضاً محك في الطبقـة البطانيـة المكونة للشعيرات الدموية في بعض الكبيبات الولية وحدوث نزيف داخل الفصيصات ولية وحوث الجسم خلايا الكبة التي أظهرة الفحوصات أيضاً محك فـي الطبقـة المانيـة المكونة للشعيرات الدموية في بعض الكبيبات التي المشت وتكونت بها حويصلات دهنية وطبقة رجاجية .

أما الدراسة الهستوكيميائية فقد أظهرت نقص في محتوى الكربوهيدرات والبروتينات وذلك بعد معاملة الفئران بجل وتاميد أحادي الصوديوم. وفيما يتعلق بالدراسات الدقيقة بواسطة الميكروسكوب الإلكتروني ، فقد أظهرت تدمير واضح في عضيّات الخلايا وتغيير في الـشكل العام للأنوية المكونة لها.

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ABSTRACT

The present investigation demonstrated the effect of oral administration of two different doses of monosodium glutamate (MSG): 2 mg/g/b.w. and 3mg/g/b.w. for 21 and 45 days respectively on the histological structure of the kidney tissues of adult male albino rats. The histopathological observation of the kidney tissues after the administration of 2mg/g/b.w., showed partial loss of brush border of proximal convoluted tubules and necrotic lesions in the epithelial lining of renal tubules. Numerous inflammatory cells infiltrated in the interstitial tissues and pyknotic, karryolysed nuclei in some tubules were also observed. Severe histopathological alterations were recorded after treatment of rats with 3mg/g/b.w. of MSG. These changes included distention of glomerular capillaries with fat globules, distortion of renal architecture in most of the cells, lobulation in some glomerular tufts with haemorhogic lesions, shrunken glomeruli which invaginated by fatty globules and diffused hyaline thickening of capillary endothelium .Histochemical investigations revealed marked reduction in both carbohydrates and proteins after the Ultrastructural studies revealed destruction of treatment with MSG. cytoplasmic organelles, thickening of the basement membrane of the proximal convoluted tubules and irregularly- shaped nuclei.

Key words: Sodium glutamate, histochemistry, histopathology, Ultrastructure, kidney.

INTRODUCTION:

A common example of one of the thousands of chemicals used in our new high-tech foods is the monosodium glutamate MSG.It is the sodium salt of the amino acid, glutamic acid and a form of glutamate.It is added to the food either as a purified monosodium salt or as a component of a mixture of amino acids and small peptides resulting from the acid or enzymatic hydrolysis of proteins, (Schwartz 2004).When it is added to this food in relatively small quantities, the palatability of those foods is increased (Yamagu ,1998). This is the substantial evidence that the sensory basis for this effect is that MSG stimulates the sense of taste (Kawamura et al., 1987).

A study conducted by (Halpern, 1997) has provided a convincing evidence that the taste quality elicited by (MSG) and related substances such as inositol monophosphate is unique. That is, it is not some combination of sweet, sour, salty and bitter, the presumed other primary taste qualities(Ohguro, et al. 2002).

Glutamate is absorbed from the gut by an active transport system specific for amino acids. This process is saturable, can be competitively inhibited, and is dependent on sodium ion concentration (Schultz et al., 1970). During intestinal absorption, a large proportion of glutamic acid is transaminated and consequently alanine levels in portal blood are elevated. If large amounts of glutamate are ingested, portal glutamate levels will increase (Stegink, 1984). This elevation results in an increase hepatic metabolism of glutamate, leading to release of glucose, lactate, glutamine, and other amino acids, into systemic circulation (Stegink, 1984).

Numerous studies have been conducted on the physiological role of MSG. Berry et al., (1974) found that it caused an enlargement of the liver and an increase of serum albumin and decrease in serum globulin. Machol, et al., (1999) found that it changed several endocrine functions in neonatally treated rats. Larsen et al.,(1994) found that it altered the activity and sensitivity of rat hypothalamo- pituitary- adrenocortical axis . Osfor-Mmh, et al., (1997) indicated that kidney ,liver ,brain ,and heart weight were significantly increased in rats treated with MSG. Miskowiak et el., (1999) found that injection of monosodium glutamate (4mg/g b.w.) to the rats resulted in a decrease of the number of Graafian follicles and lowered the thickness of endometrial controls. MSG induced alterations in metabolic rate of glucose utilization and decreased antioxidant defenses(Diniz-Ys,et al., 2004).

There uses have become controversial because of some reports of adverse reaction in people who have eaten foods that contained MSG. Studies conducted by Schwartz (2004) have shown that the body uses glutamate as a nerve impulse transmitters in the brain, and that there are glutamate responsive tissues in other parts of the body. The MSG treated rats showed a severe growth retardation as a consequence of the impairment of r GH anabolic effects (Sakai et el., 2004). Description of the pathological effects of MSG on organs of the rats were lacking. So, the present study aims to highlight on the histopathological alterations of the kidneys of albino rats treated with MSG.

MATERIAL AND METHODS:

Sixty adult male albino rats (Rattus norvigicus) weighing 100 ± 10 g. were used in the present study. They were obtained from Helwan laboratory farms for the Egyptian Organization for Vaccine and Biologic Preparations. The rats were kept in the laboratory for one week before the experimental work and maintained on a standard diet and water available ad libitum.

The rats were equally divided into three groups and orally treated as follows:

Group(1): The control group in which rats were administrated with 0.9% NaCL day after day for 45 days.

Group(2): The experimental group in which rats were administrated orally with the therapeutic dose of monosodium glutamate (2mg/gm body weight) dissolved in 0.9% NaCL day after day for 21 days.

Group (3): The treatment group in which rats were administrated orally with the therapeutic dose of monosodium glutamate (3mg/gm body weight) dissolved in 0.9% NaCl day after day for 45 days.

Histological and Histochemical Preparations:

The kidneys from the control and experimental groups were rapidly excised after the previously mentioned duration, cut into small pieces and dropped in Bouins fluid in which they were kept for appropriate time. After fixation, they were subjected to the normal procedure for paraffin embedding. Sections were cut at the thickness of 5 microns and stained with haematoxyline-eosin(Drury and Wallington, 1976). Periodic Acid Schiff technique (PAS)(Hotchkiss, 1948), was used for the studying of general

carbohydrate and bromophenol blue technique) was used for the demonstrating of total proteins(Mazia et al.,1953).

Transmission Electron Microscopy Studies:

Small pieces (1 mm) of treated tissues were cut and fixed in 3% glutaraldehyde (PH7.4) in phosphate buffer and post fixed in 2% osmium tetroxide in phosphate buffer. Following fixation, tissues were dehydrated at increasing concentrations of ethanol. They were then embedded in araldyte resin. ultrathin sections were cut using an ultratome. Ultrathin sections were stained by uranyl acetate saturated in 70% ethanol, and lead citrate (*Reynolds,1963*). Tissue sections were evaluated using a JEOL transmission electron microscope JEM-1200. Ex, Japan.

RESULTS:

Histological Observations:

Group I: kidney sections of control rats showed normal histological structures of the glomruli, and renal tubules in the cortical (Fig.1) and medullary portions(Fig.2).

1. Histopathological Observations:

Group II : Rats treated with a dose of 2 mg/gm b.w. of MSG for 21 days showed variable pathological changes in glomeruli and some parts of the urinary tubules. Such changes exhibited an existence of hyperemia with swelling in the lining epithelium of the glomerulus associated with a priglomerular focal area. Also, they were represented by dramatic renal injury with tubular cell swelling, loss of brush border of proximal convoluted tubules as well as necrotic lesions (Fig.3).Dilatation and hyperemia in the intertubular cortical blood vessels were seen clearly in figure (4).

Swelling in the lining epithelium of the renal tubules with narrow lumen and the presence of inflammatory cellular infiltration, mainly composed of lymphocytes and monocytes in interstitial tissues were noticed in figure (5). Moreover, figure (6) exibited vascular degeneration in the epithelial cells lining the renal tubules at the cortical zone with pyknotic and karyolysed nuclei .The pathogenic observations in figure (7,9) showed hyperemic capillaries, focal hemorrhage between the degenerated renal tubules at the

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corticomedullary portion as well as Karyolysis in the epithelial lining of the urinary tubules with an infiltration of chronic inflammatory cells that shown which tend to concentrate around tubules (Fig.8).

Group III: The histopathological changes in rats received a dose of 3 mg/gm/b.w of MSG for 45 days in alternate days manifested more intensive deterioration and damage than those observed in group II. Figures (9,10) revealed distension of glomerular capillaries with fat globules .There was an increase in the incidence of marked severe vascular degenerative changes in the lining epithelial cells of the renal tubules at the cortical portion and distortion in the renal architecture (fig.11). On the other hand, figure (12) showed shrunken of many glomeruli invaginated by fatty globules, diffused hyaline and thickening of capillary endothelium. In addition, there was a focal mononuclear leukocytes inflammatory cells that infiltrate between the tubules at the corticomedullary portion (Fig.13).

With the progress of time, more degenerative changes took place in the renal tubular epithelium cell debris in their lumens (Fig. 14). Also, lobulations in some glomerular tufts with haemorrhagic lesions were seen in figure (15). Necrosis of other tubules lining cells , focal mononuclear leucocytes , inflammatory cells infiltrating renal tubules , focal haemorrhagic area in between the renal tubules and chronic inflammation replaced urinary tubules were evident in all rats treated with MSG in this group (Figs. 16, 17).

The most severe changes were in the proximal convoluted tubules which consisted of cytoplasmic vacuolation of tubular epithelial cells with swelling and detachment of them. With the progress of time, interstitial edema, more extensive necrosis of tubular epithelial cells, and dilated tubules with accumulation of eosinophilic homogenous material in tubular lumina (Fig.17).

Histochemicals Observations:

1.General carbohydrates:

A considerable amount of carbohydrates in the cytoplasm of kidney cells of control rats was noticed by PAS-technique, which gave a red or magenta colour (Fig 18). The nuclei, however, appeared entirely PAS-negative staining, indicating absolute lack of carbohydrates.

Treating rats with 2 mg/gm/b.w of MSG for 21 days, caused a decrease of total carbohydrates in the kidney cells (Fig.19). A marked reduction of total

carbohydrates was observed after 45 days of treatment with 3 mg/gm/b.w MSG (Fig. 20).

2. Total proteins:

Total proteins were demonstrated in epithelial cells lining the renal tubules of control rats as deeply blue stained diffuse granules homogenously through both the cytoplasm and nuclei (Fig 21). Their nuclear envelopes and nucleoli as well as some chromatin elements were also positively stained (Fig 21).

Treatment of rats with 2 mg/gm/b.w MSG for 21 days induced a slight decrease in the protein content of the urinary tubules (Fig 22). More reduction in proteins was manifested in the cells treated for 45 days, where the proteinic granules were clearly reduced in amount and stainability (Fig 23).

3.Ultrastructural Observations:

The cells of the proximal convoluted tubules of control rats showed long rod-like mitochondria oriented parallel to the cell axis. They have long microvilli forming the brush border appeared and large rounded nuclei (Fig. 24).

The Ultrastructural examinations revealed changes of the cells of the renal tubules after administration of 2 mg/gm /b.w of MSG. These comprised intracellular vacuolations , destruction of cytoplasmic organelles, splitting and thickening of basement membrane (Fig. 25). Additionally, other cells of proximal tubules showed irregularly- shaped of nuclei with loose chromatin materials and invagination of the nuclei membrane (Fig.26).

DISCUSSION :

The present study indicated that MSG induced marked histopathological alterations in the kidney tissues of rats such as tissue impairment ,swelling of the lining epithelium of glomeruli, injured brush border of proximal convoluted tubules, necrotic lesions of the urinary tubules and focal hemorrhage between the degenerative renal tubules. Similar results, have been reported by others (Aughey et al., 1984; kjellstrom, 1986; Mitsumari et al., 1998); Inkielewicz and Krechniak (2003).

The mechanism of swelling starts as a decrease in O2 levels which causes a drop in aerobic respiration. To maintain ATP levels, the cells must rely more on glycolysis. Glycolysis leads to lactic acid builds up, which causes the intracellular pH to drop. An acidic environment in the cell causes dysfunction of the Na⁺/K⁺ ATPas and consequent cell swelling due to an influx of Na⁺ and H₂o.Persistent ischemia can lead to ca⁺⁺ influx mitochondrial and lysosomal damage, and membrane damage (Lieberthat et al., 1998).

In the present investigation, many renal tubules of the rat kidneys showed marked degenerative lesions under the effect of MSG. This is justifiable since the renal tubules are particularly sensitive to toxic influences, in part because they have high oxygen consumption and vulnerable enzyme systems, and in part because they have complicated transport mechanisms that may be used for transport of toxins and may be damaged by such toxins. Also the tubules come in contact with toxic chemicals during their excretion and elimination by the kidneys (Tisher and Brenner, 1989). Such degenerative changes were markedly pronounced in the proximal convoluted tubules. These findings reinforce those of Koechel et al., (1984) and Damjanov (1996), who found that many chemicals had a direct nephrotoxic action and exerted their effects principally on the proximal convoluted tubules.

The presence of necrosis may be related to the depletion of ATP, which finally leads to the death of the cells (Shimizu et al., 1996). Renal medullary necrosis occurs as a primary manifestation of renal disease. The mechanism of which is poorly understood, but it seems to involve a vascular change. Also, prostaglandin synthetase is found in the kidney, primarily in the medulla, and inhibition of this enzyme resulted in decreased production of prostaglandin E2 (PGE2) and loss of its vasodilatory effect on juxtamedullary arterioles. (Date and Shastry, 1982).

De Camargo and Merzel,(1980) observed that mice fed with 10 and 500 ppm NaF for 3 months had shrunken kidneys, degeneration of tubular cells, and dilatation in the convoluted tubules. Similar changes were seen in the present investigation.

One possible mechanism for the tubular lesions was the direct toxic effect on the cell function (Alden and Frith, 1992). Damage to the brush border and leakage of alkaline phosphates (ALP) and gammaglutamyle transferase (GGT) enzymes , which are associated with the brush border of the renal tubules, as a result of toxin binding to the brush border and considered as an early marker of toxic tubular insult (Edelstein, et al., (1995); Davies et al.,(1995); Porter , (1994); Fadel ,and Larsen, (1994). Other possible mechanisms for the tubular lesions may involve reactive intermediates or oxidative stress, or both (Alden and Frith 1992). Biologically reactive intermediates are electron-deficient compounds (electrophiles) that bind to cellular electron-rich compounds, such as proteins and lipids (Goldstein and Schnellmann, 1995). Mixed-function oxidases catalyze the formation of there toxic metabolites. Reactive intermediates bind covalently to critical cellular macromolecules and interfere with normal biologic activity. Oxidative stress is induced by increasing production of reactive oxygen specie (ROS), such as superoxide anion, hydrogen peroxide and hydroxyl radicals (Goldstein, et al., 1995). ROS can induce lipid peroxidation, inactivate cellular enzymes, depolymerize polysaccharides, and induce deoxyribonucleic acid breaks and chromosome breakage.

Superoxide dismutase (SOD) is a naturally occurring intracellular enzyme that catalyzes the breakdown of superoxide radicals (McCord, 1993). Ischemia leads not only to an increase in superoxide production, but also, to a rapid depletion of SOD (singh et al.; 1993 and Davies et al.; 1995).

The detection of Lymphocyte inflammatory cells in the present study indicated the production of chronic inflammatory disease under the effect of MSG. This result agreed with Ashry et al., (1990) who demonstrated chronic active cells accompanied by inflammatory cells in the hepatocytes after administration of codeine.

The results showed that treated rats with MSG caused a depletion of carbohydrates in the cytoplasm of renal tubules. This result was in correspondence with other studies reported by Sakr et al.; (2003) due to the treatment of gibberellin to the rats, and Abdeen et al.,(1994) and Sakr et al.,(2002a) duo to the use of a variety of animals under different pathological conditions.

Disturbances in carbohydrate metabolism were also observed in a variety of animals under the effect of different insecticides and were suggested to be achieved through modifying the activities of the enzymes of glycolytic pathway. TCA cycle, glucogenesis and the oxidation of phosphorylation (Kacew and Singhal, 1973 and Shakoori et al., 1988).

The present study also revealed that treatment rats with MSG induced marked decrease in protein contents of the studied cells. This result consistent with other studies reported by Mehadevaswami et al., (2001) due to the use of different compounds.

The reduction of protein contents observed in this study may be attributed partially to the decrease of hepatic protein synthesis due to the hyperactivity of hydrolytic enzymes (Sivaprasada et al., 1983). Moreover, the decrease occurred in proteins is likely to be a consequence of the damage produced by MSG in the rough endoplasmic reticulum and Golgi apparatus as reported by (Elewa et al., (1999), El-Beih et al., (1987)and Berlin (1967). In addition, Palla et al., (1987), postulated that in many kidney diseases, the permeability of the glomerular capillaries is increased leading to increased levels of excreted proteins. They added that any lesions produced in the kidney tubules will eventually cause dysfunction in the transport mechanism to and from the renal epithelium.

The electron microscopic examination revealed destruction in the cytoplasmic organelles including lysosomes, endoplasmic reticulum, mitochondria and Golgi apparatus. These alterations could be due to the cytotoxic effects of MSG. Thomas (1988) stated that Golgi apparatus is responsible for the packaging of hydrolytic enzymes involved in the formation of secretary products. The destruction of Golgi apparatus leads to destruction of lysosomes and this consequently leads to an increase in secretion of hydrolytic enzymes which may be responsible for the lysis of cytoplasmic organelles.

Finally, the present study reveals toxic effects of MSG on the kidney during the use of this drug. Therefore, more researches must be done on other organs of the body to highlight its effects on these organs.

REFERENCES:

1. Abdeen, AM.; Amer TA.; EL-Habibi, EM.; and Kamal EM.(1994): Histological and histochemicals studies on the effect of fenvalrate insecticide on some organs of Albino mice. J Union Arab Biol 2A, 129-66

2. Alden, CL, and Frith CH. (1992): Urinary System. In: Handbook of Toxicologic Pathology, ed. Hashek WM and Rousseaux CG, Ist ed., pp 316-379, Academic Press, San Diego, CA.

3. Ashry, M.; Wahba; S. and Abdel-Mageid, S. (1990): Histological and histochemicals changes in response to the administration and withdrawal of codeine on liver of rat. Egypt. J. Histol., 13: 3-12..

4. Aughey, E.; Feli, G.S.; Scott, R, and Black M. (1984): Histopathology of early effects of oral cadmium in the rat kidney, Environment and Health Perspectives 54, 153-161.

5. Berlin, J.D. (1967): .The localization of acid mucopolysaccharides in Golgi complex of intestinal goblet cells. J. Cell Biol., 32: 670-766.

6. Berryry, H.K; Butcher, R.E; Elliot, L.A; Brunnr, R.L. (1974) the effect of nonosodium glutamate on the early bioxhemical and behavioural development of the RAT: Dev-Psyhobiol. 1974 Mar; 7(2): 165-73

7. Damjanov, I. (1996): Histopathology: A Color Atlas and Textbook. pp. 257-287. Williams and Wilkins. A Waverly Company. Baltimore. Philadelphia and London.

8. Date, A and Shastry, J. C.M. (1982): Renal ultrastructure in acute tubular necrosis following Russell's viper envenomation. J. Pathol., 137:225-241.

9. Davies, S.; Reichardi; Pascal SY.; Vaughan D.; Reussell GI (1995): Differential effect of ischemia-reperfusion injury on anti-oxidant enzyme activity in the rat kidney. Exp Nephrol 3:348-354.

10. De Camargo , AM.; Merzel J.(1980); Histological and Histochemical appearance of lives and kidneys of rats after long-term treatment with different concentrations of sodium fluoride in drinking water. Acta Anat; 108: 288-94

11. Diniz, YS,; Fernandes-Aah; Campos-Ke ; Mani-F (2004): Toxicity of hypercaloric diet and monosodium glutamate : Oxidative stress and metabolic shifting in hepatic tissue . Food-and- chemical – Toxicology , 42:2, 313-319

12. Drury R.A.B. And Wallington E.A. (1976): Carleton's Histological Technique. Oxford University Press, New York, 4th ed. P. 129.

13. Edelstein CL, Ling H, Gengaro Pe, Nemenoff Ra, Bahr BA,(1995): Effect of glycine on prelethal and postlethal increases in rats blood. Res. Vet. Sci. 58:180-188

14. El-Beih, Z.M.; Amer, M.A. and Elewa, F.H. (1987): Histochemical observations on the mucopolysaccharides in duodenal mucosa of normal and insecticide-treated guinea pigs Bull. Fac. Sci., Cairo Univ., 55: 65-75.

15. Elewa, F.H.; GABRY, M.S.; AND IBRAHIM, M.A. (1999): Ultrastructural changes produced by diclofenac sodium in the liver and duodenal epithelial cell of the guinea pig. Egypt. J. Zool., 33: in press.

16. Fadel , A.A, And Larsen H.A. (1996): Gentamicin-induced nephrotoxiois in lambs. Res. Vet. Sci. 61:187-192

17. Goldstein, R.S. and Schellmann R.G. (1995): Toxic responses of the kidney. In: casarett and Doull's Toxicology. The Basic Science Of Poisons, ed., Klaassen CD, 5 th ed., pp 417- 442, McGraw-Hill companies Inc., New York, NY.

18. Halpern, B.P. (1997): Psychophysics of taste. Beauchamp G.R. Bartoshuk l. eds. Tasting & Smelling (77-123) Academic press San Diego, CA.

19. Hotchkiss, R.D. (1948): A microchemical reaction resulting in the staining of polysaccharidd structure in fixed tissue preparations. Arch. Biochem., 16:131.

20. Inkielewicz, I. and Krechniak(2003): Fluoride content in soft tissues and urine of rats exposed to sodium fluoride in drinking water, fluoride vol. 36 No. 4 263-266 2003 Research Report 263

21. Kacew, S. and Singhal, L. (1973): Adaptive response of carbohydrate metabolism to oral administration of p.p. 1, 1-Trie chlorophonly) ethane. Biochem. Pharmacol., 22:4 7-57)

22. Kawamura, Y.; Kare, M.R. EDS. Umami (1987): A Basic Faste Marcel Dekker. New Yourie, N Y.

23. Kjellstrom, T., (1986): Renal effects in cadmium and health: A Toxicology and Epidemiological Appraisal, Vol. 2, Friberg, L., Elinder, C.G., Kjellstrom, T. and Norgderg, G.F eds, pp. 21-109, CRC Press, Boca Raton.

24. Koechel, D. A.; Bretz, N.S.; Sanzenbacher , R.L. and Tarloff, J. B. (1984): The pentobarbital anesthetized dog: An animal model for assessing chemically-induced changes in renal function and Ultrastructure. Am.J. Vet. Res. 45(12): 2565-2573.

25. Larsen, P.J.; Mikkelsen, J.D.; Jessop, D.; Lightman, S.L. and Chwoday, H.S. (1994): Neonatal monosodium glutamate treatment alters both the activity and the sensitivity of the rat hypothalamo-pituitary-adrenocotical axis. J. Endocrino. 141:497-503

26. Lieberthat, W.; Menza S.A.; Levine J.S. (1998): Graded ATP depletion can cause necrosis or apoptosis of cultured mouse proximal tubular cells. Am J Physiol 274: F315-F327.

27. Machol, Jeove, D.; Zarad M.; and Fickov S. (1999): Effect of Early postnatal monosodium treatment on adrenal cortex function. Institute of Experimental Endocrinology, SAS, Bratislava, Slovakia.

28. Mazia , D.; Bewer P.A.; and Affest M.; (1953): The cytochemical staining and measurements of protein with mercuric bromophenol blue. Biol Bull 104,57-67.

29. McCORD , JM. (1993). Human disease, free radicals and the oxidant/antioxidant balance. Clin Biochem 26:351-357.

30. Mehadevaswami, M. P.; Jardarankunti, M.B.; Hiremath U.C B.B. Kaliwal, (2001): Effect of mancozeb on ovarian copensatoy hyprtrophy and biochemical constituents in hemicastrated albino rat Reorod. Toxicol., 14:127-137.

31. Miskowiak , B.; Kesa B.; Limanowski A.; Pantayka M.; Filipiak B.; Folia Monpho (Woasz). (1999): Long-term effect of neonatal (MSG) treatment on reproductive system of the level rats. 58 (2): 105-13

32. Mitsumari , K.; Shibutani S.; Sato S., Ondoera H.; Nacagawa J.; Hayashi Y.; and Ando M.; (1998): Relationship between the development of hepatorenal toxicity and cadmium accumulation in rats. Archives of Toxicology 72,545-552

33. Ohguro , H.; Katsushima H.; Maruyama , I.; Maed T.; Yanagihashi, S.; Metoki , T.; and Nakazawa M (2002): A high dietary intake of sodium glutamate as a flavoring (a jinomoto) causes gross changes in retinal morphology and function Exp Eye Res. 75 (3): 307-15.

34. Osfor-Mmh; El-Desouky, SA.; El-Leithy NA.; (1997): Effect of dietary intake of monosodium glutamate on some nutritional and biochemical traits in albino rats, Egyptian-Journal of comparative-pathology and Clinal pathology . 10:2, 131-139

35. Palla , R.; Patrenos Terg G.; Galigane, R; Brtell, A.; Romono, M.; Alessandr, M. and Bartalla, A. (1987): Comparative effects of gentamicin, amikacin and dactimicin on excretion of acety1 beta-d-glucosamidase (Nag) and kidney histological pattern.

36. Portr , GA.; (1994): Urinary biomarkers and nephrotoxicity. Miner Electrolyte Metab 20:181-186.

37. Reynolds, E.A.(1963): The use of PH as an electron opaque stain in electron microscopy. J.Cell Biol. 17 : 208.

38. Sakai , T.; Miki F.; Warrishi M.; Yamamoto S. (2004): Comparative study of Zink, copper, Manganese & brain concentrations in organs of Zink-deficient Rats and rats treated neonatally with MSG. Biol Trace Elem Res; 97 (2):163-82.

39. Sakr , S.; Okdah A.; and El-Abed F. (2003): Gibberellin A3 Induced Histological and Histochemical Alterations in the liver of Albino Rats, SienceAsia 29 (2003): 327-331.

40. Sakr , SA. El-Messedy, FA and Abdel-Samei Ha. (2002a): Histopathological and histochemicals effects of gibberellin A3 on the kidney of albino rats. J Egypt Grrm Soc Zool 38.1-10

41. Schultz, S.G.; Yu-Tu, L., Alvafez, O.O.; and Curran, P.F. (1970): Dicarboxylic amino aid influx across brush border of rabbit ileum. J. Gen. Physiol., 56, 621-639

42. Schwartz, J.R. (2004): In bad taste, the MSG "syndrome" MSG. the 5th Annual Conference of the Weston A. Price Foundation.

43. Shakoori, A. R.; Ali; S. S. and Saleem, M. A. (1988): Effect of six months feeding of cypermethrin on the blood and liver of albino rats. J. Biochem. Toxicol, 3:59-72

44. Shimizu , S.; Eguchi Y.; Kamiike W.; Waguri S.; Uchiyama Y.; Matsuda , H.; Tsujimoto Y.; (1996): Retardation of chemical hypoxiainduced necrotic cell death by Bcl-2 and ICE inhibitors: Possible involvement of common mediators in apoptotic and necrotic signal transductions. Oncogene 12: 2045-2050.

45. Singh , I.; Gulati S.; Orak Jk, and Singh AK.; (1993): Expression of antioxidant enzymes in rat kidney during ischemia-reperfusion injury. Mol Cell Biochem 125:97-104.

46. Sivaprasada , K.; Sombasiva; K. R., and Ramana, K. V. (1983): Effect of parathion on tissue ionic changes fish channa punctatus Geobios. Jodhpur, 10:60-62

47. Stegink , L.D. (1984): Aspartate and glutamate metabolism. Aspartame: physiology and biochemistry, pp. 47-76.

48. Thomas , S (1988): Text atlas of Histology W.B. Saundrs company , Philadelphia.

49. Tisher, C.C. and Brenner. B. M. (1989): Renal Pathology with Clinical and Functional Correlation. Volume (1) J. B. Lippincott company. Philadelphia.

50. Yamagu Chi , S. (1998): Basic properties of umami and its effects on food flame. Food Rev. Int. 14: 139-176.

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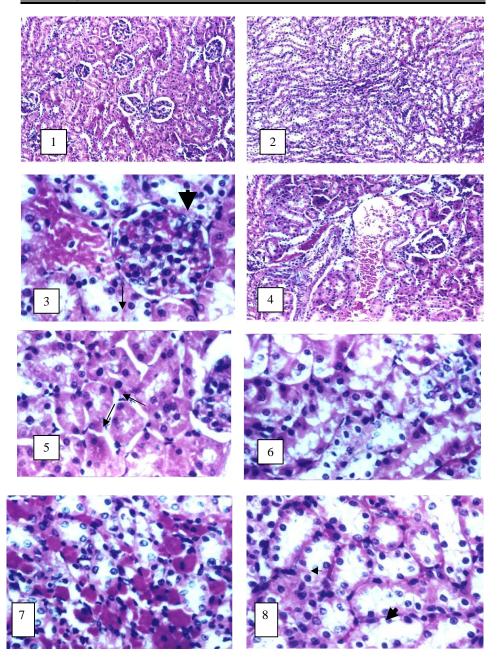
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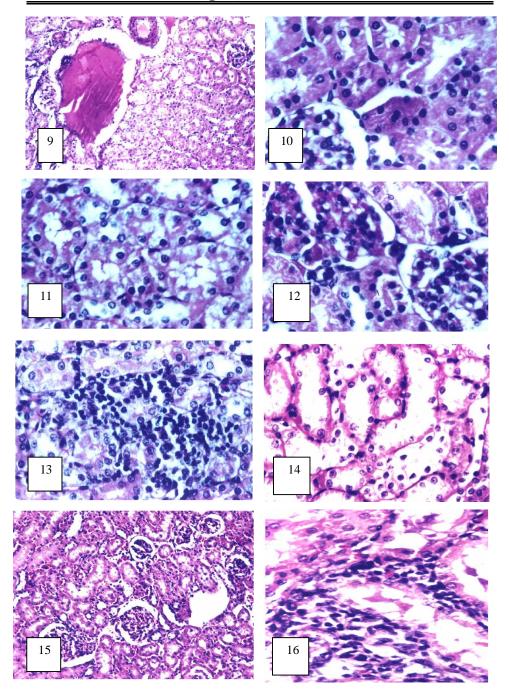
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